

TECHNICAL MANUAL

Lumit[®] IFN- β (Human) Immunoassay

Instructions for Use of Products
W1810, W1811 and W1812

Lumit[®] IFN- β (Human) Immunoassay

All technical literature is available at: www.promega.com/protocols/
 Visit the website to verify that you are using the most current version of this Technical Manual.
 Email Promega Technical Services if you have questions on use of this system: techserv@promega.com

| | |
|---|----|
| 1. Description | 2 |
| 2. Product Components and Storage Conditions | 3 |
| 3. Before You Begin | 4 |
| 4. Direct (No-Transfer) Protocol for Cultured Cells | 6 |
| 4.A. Cell Plating and Treatment..... | 6 |
| 4.B. Preparing Human IFN- β Standard Dilution | 7 |
| 4.C. Adding 5X Anti-hIFN- β Antibody Mixture to Assay Wells..... | 8 |
| 4.D. Adding Lumit [®] Detection Reagent B to Assay Wells | 9 |
| 5. Optional Sample Transfer Protocol | 10 |
| 5.A. Cell Plating and Treatment..... | 10 |
| 5.B. Preparing Human IFN- β Standard Dilutions | 11 |
| 5.C. Adding 2X Anti-hIFN- β Antibody Mixture to Sample Wells..... | 12 |
| 5.D. Adding Lumit [®] Detection Reagent B to Sample Wells..... | 13 |
| 6. Calculating Results | 14 |
| 7. Representative Data | 14 |
| 8. Troubleshooting | 16 |
| 9. Appendix..... | 17 |
| 9.A. References | 17 |
| 9.B. Related Products..... | 18 |

1. Description

The Lumit[®] IFN- β (Human) Immunoassay^(a,b,c) is a homogeneous, bioluminescent assay for detecting interferon-beta (IFN- β) released from cells without the need for sample transfers or wash steps.

The Lumit[®] IFN- β (Human) Immunoassay has been developed for use with cell culture samples. Lumit[®] reagents can be dispensed directly into microplate wells containing cells and culture medium. Alternatively, medium from cell wells can be transferred to a separate plate for analysis. Assay performance with sample types other than cell culture medium must be determined by the user.

The Lumit[®] IFN- β (Human) Immunoassay is based on NanoLuc[®] Binary Technology (NanoBiT). NanoBiT is a luminescent structural complementation system designed for biomolecular interaction studies (1,2). The system is composed of two subunits, Large BiT (LgBiT; 18kDa) and Small BiT (SmBiT; 11 amino acid peptide), that have been optimized for stability and minimal spontaneous association. In this assay, a sample is incubated with a pair of anti-human IFN- β monoclonal antibodies covalently labeled with SmBiT or LgBiT. When the labeled antibodies recognize and bind to released IFN- β , the complementary LgBiT and SmBiT are brought into proximity, thereby reconstituting the NanoBiT[®] enzyme and generating luminescence in the presence of the Lumit[®] substrate. Luminescence generated is directly proportional to the amount of analyte present in the sample.

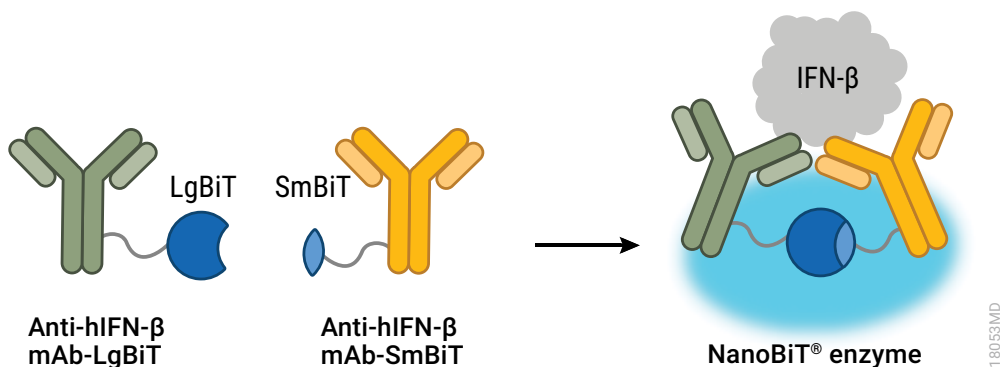


Figure 1. Assay principle. Primary monoclonal antibodies to human IFN- β are labeled with SmBiT and LgBiT. In the presence of IFN- β , SmBiT and LgBiT are brought into close proximity, forming the NanoBiT[®] enzyme. When Lumit[®] detection reagent B is added, a bright luminescent signal is generated.

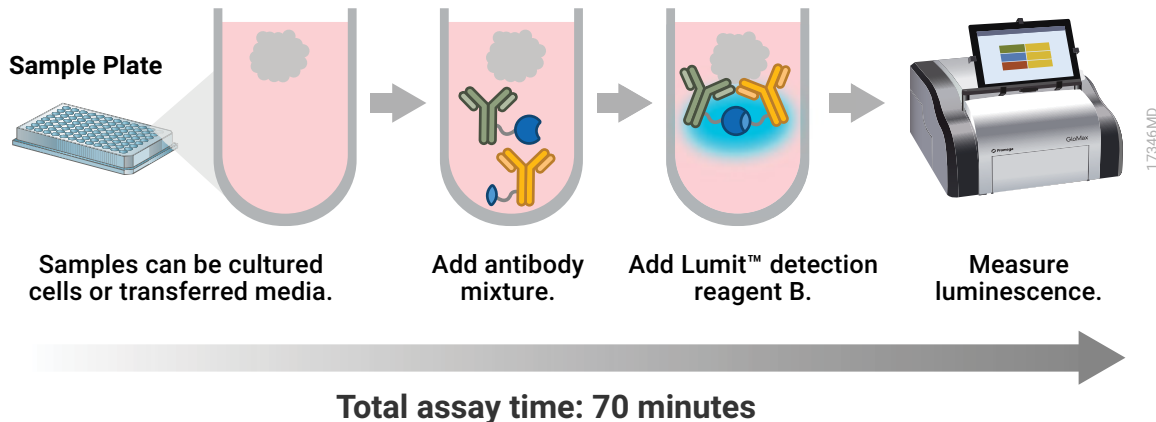


Figure 2. Assay protocol. The Lumit® IFN-β (Human) Immunoassay is performed directly on cells in culture or medium transferred from the cell culture plate to a new assay plate. The Lumit® Immunoassay protocol does not require wash steps and can be completed in 70 minutes. Antibody mixture consists of Anti-hIFN-β mAb-SmBiT and Anti-hIFN-β mAb-LgBiT.

2. Product Components and Storage Conditions

| PRODUCT | SIZE | CAT. # |
|---|-------------------|--------------|
| Lumit® IFN-β (Human) Immunoassay | 100 assays | W1810 |

Sufficient for 100 assays in 96-well plates; adjust assay volume for alternate plate sizes. Includes:

- 15µl Anti-hIFN-β mAb-SmBiT, 1,000X
- 15µl Anti-hIFN-β mAb-LgBiT, 1,000X
- 25µl Human IFN-β Standard
- 160µl Lumit® Detection Substrate B
- 3.2ml Lumit® Detection Buffer B

| PRODUCT | SIZE | CAT. # |
|---|---------------------|--------------|
| Lumit® IFN-β (Human) Immunoassay | 1,000 assays | W1811 |

Sufficient for 1,000 assays in 96-well plates; adjust assay volume for alternate plate sizes. Includes:

- 150µl Anti-hIFN-β mAb-SmBiT, 1,000X
- 150µl Anti-hIFN-β mAb-LgBiT, 1,000X
- 25µl Human IFN-β Standard
- 1.25ml Lumit® Detection Substrate B
- 25ml Lumit® Detection Buffer B

2. Product Components and Storage Conditions (continued)

| PRODUCT | SIZE | CAT. # |
|--|-----------------------|--------------|
| Lumit[®] IFN-β (Human) Immunoassay | 5 × 100 assays | W1812 |

Sufficient for 500 assays in 96-well plates; adjust assay volume for alternate plate sizes. Includes:

- 5 × 15μl Anti-hIFN-β mAb-SmBiT, 1000X
- 5 × 15μl Anti-hIFN-β mAb-LgBiT, 1000X
- 25μl Human IFN-β Standard
- 5 × 160μl Lumit[®] Detection Substrate B
- 5 × 3.2ml Lumit[®] Detection Buffer B

Storage Conditions: Store complete kit at less than -65°C upon receipt. Alternatively, store the Human IFN-β Standard at less than -65°C and all other components at -30°C to -10°C. After thawing, Human IFN-β Standard can be stored for up to 1 month at 4°C. If storing the Human IFN-β Standard for more than 1 month after thawing, dispense into aliquots and store at less than -65°C. After thawing, store the Anti-hIFN-β mAb-SmBiT and Anti-hIFN-β mAb-LgBiT at -30°C to -10°C. After thawing, store Lumit[®] Detection Buffer B at room temperature. Store Lumit[®] Detection Substrate B protected from light.

3. Before You Begin

There are two protocols for measuring human IFN-β (hIFN-β). The direct protocol typically achieves higher sensitivity than the optional sample transfer protocol and requires less sample manipulation. The optional sample transfer protocol provides flexibility for same well sampling during treatment exposure time courses and split-sample analysis for assessment of multiple cytokine levels from the same sample.

Direct (No-Transfer) Protocol for Cultured Cells (Section 4): Measure human IFN-β directly in cell culture wells. Add 20μl of a 5X antibody mixture to 80μl of cells or IFN-β standard dilutions in culture medium and incubate for 60 minutes. Following incubation, add 25μl of Lumit[®] detection reagent B and record luminescence.

Optional Sample Transfer Protocol (Section 5): Measure human IFN-β in medium samples transferred from treated cell wells. Transfer 50μl of culture medium from cell wells to a separate assay plate. Add 50μl of a 2X antibody mixture to 50μl of transferred sample or standard dilutions and incubate for 60 minutes. Following incubation, add 25μl of Lumit[®] detection reagent B and record luminescence.

Note: Assay volumes are scalable and can be adjusted based on sample sizes. The protocols in Sections 4 and 5 list common volumes for 96- and 384-well plates. Other volumes can be used as long as you maintain the recommended antibody and detection reagent final concentrations. We recommend using standard tissue culture medium supplemented with 5–10% fetal bovine serum (FBS). Lesser concentrations of FBS may produce higher background (nonanalyte-mediated antibody pairing) and reduced assay sensitivity. When using serum-free, albumin-free medium, assay background may be reduced and sensitivity improved by preparing the assay antibody mixture in medium supplemented with bovine serum albumin (BSA) to deliver 0.05% BSA final after addition to sample. (Using significantly higher albumin concentrations may further compromise assay performance.) Using medium without phenol red may increase assay sensitivity and reduce inner-filter effects from luminescence quenching.

Reagent Preparation and Storage

Prepare the Human IFN- β Standard dilution series (Section 4.B or 5.B), the Anti-hIFN- β antibody mixture (Section 4.C or 5.C) and Lumit[®] detection reagent B (Section 4.D or 5.D) on the day of use. Do **not** reuse Human IFN- β Standard dilutions, the Anti-hIFN- β antibody mixture or the Lumit[®] detection reagent B. To avoid reagent contamination, use only fresh aerosol filter tips or cotton-plugged, sterile pipettes when withdrawing volumes from reagent stock components.



Use personal protective equipment and follow your institution's safety guidelines and disposal requirements when working with biohazardous materials such as cells and cell culture reagents.

Plate Map

| | IFN- β Standard Curve (pg/ml)* | | Test Samples | | | | | | | | | |
|----------|--------------------------------------|--------|--------------|---|---|---|---|---|---|----|----|----|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
| A | 25,000 | 25,000 | | | | | | | | | | |
| B | 7,500 | 7,500 | | | | | | | | | | |
| C | 2,250 | 2,250 | | | | | | | | | | |
| D | 675 | 675 | | | | | | | | | | |
| E | 203 | 203 | | | | | | | | | | |
| F | 60.8 | 60.8 | | | | | | | | | | |
| G | 18.2 | 18.2 | | | | | | | | | | |
| H | 0 | 0 | | | | | | | | | | |

***Note:** The indicated Human IFN- β Standard dilution series listed above and described in the subsequent protocol directions are only a recommended dilution series.

Materials to Be Supplied by the User

- human-derived cells
- culture medium (e.g., RPMI 1640, GIBCO Cat. # 22400-089 and 10% heat-inactivated, fetal bovine serum, GIBCO Cat. # A3840001)
- white, multiwell tissue culture plates (solid white or white with clear bottom) compatible with a luminometer (e.g., 96-well Corning Cat. # 3917)
- multichannel pipette or automated pipetting station
- aerosol filter pipette tips
- dilution tubes or multichamber, dilution reservoir (e.g., Dilux D-1002)
- reagent reservoir trays (e.g., Midwest Scientific Cat. # RR25)
- plate shaker for mixing multiwell plates
- luminometer capable of reading multiwell plates (e.g., GloMax[®] Discover System, Cat. # GM3000)

4. Direct (No-Transfer) Protocol for Cultured Cells

This protocol describes how to detect IFN- β released directly in assay wells containing cells and culture medium. For quantitation purposes, a standard curve is generated using an IFN- β standard diluted in culture medium.

4.A. Cell Plating and Treatment

1. Plate cells into a 96- or 384-well, white (or white with clear bottom) tissue culture plate. Leave columns 1 and 2 empty. Allow cells to attach if using adherent cells.

Note: While the broad dynamic range of the assay offers considerable flexibility for cell number, the optimal cell number dispensed per well for a specific cell model should be empirically determined. Ensure the maximum level of cytokine released does not exceed the linear range of the detection chemistry. Within that constraint, cell number can be increased to meet the detection requirements of low-level cytokine production.

2. Treat cells by adding a volume of test agent to each well such that the total volume is as follows:

96-well plate: 80 μ l per well.

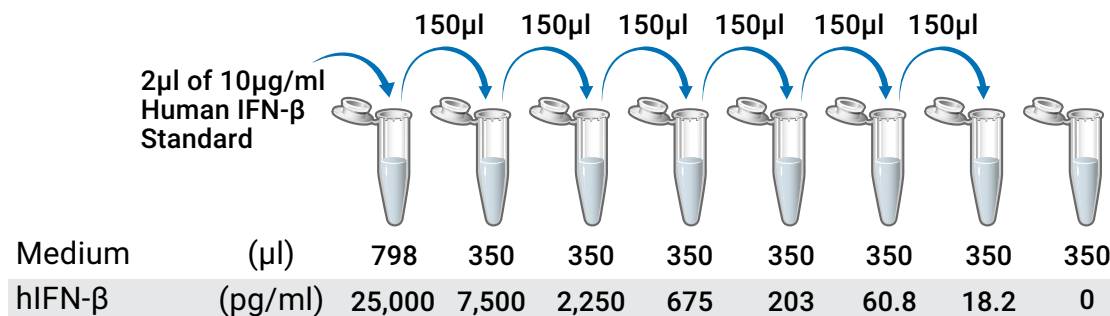
384-well plate: 20 μ l per well.

For example, if 60 μ l of cells are plated per well in a 96-well plate, add 20 μ l of 4X treatment agent in culture medium. Cells are typically treated for 6–24 hours or longer, depending on stimuli, to observe significant IFN- β release.

Optional: If manually dispensing into a 384-well assay format, after various additions, briefly centrifuge the microplate (700–900rpm for approximately 10 seconds) then briefly mix with a plate shaker.

Note: Assay sensitivity may be reduced in 384-well format compared to that in 96-well format.

4.B. Preparing Human IFN- β Standard Dilution



18054MD

Figure 3. Human IFN- β dilution series.

Shortly before completing cell treatments, prepare IFN- β dilutions in the identical culture medium used for cell samples.

Note: If using multiple cell models, each requiring a different culture medium, separate standard dilution series must be generated in each medium used in the study.

1. Thaw the Human IFN- β Standard (approximately 15 minutes at room temperature) immediately before use.
2. Briefly centrifuge the tube before opening, then mix by pipetting.
3. Prepare an initial concentration of 25,000pg/ml human IFN- β by diluting Human IFN- β Standard (10 μ g/ml) 1:400 in prewarmed cell culture medium. For example, prepare 800 μ l of 25,000pg/ml human IFN- β by adding 2 μ l of the Human IFN- β Standard stock to 798 μ l of culture medium (see Figure 3).
4. Set up seven tubes (or seven chambers in a dilution reservoir) with 350 μ l of culture medium in each.
5. Prepare 3.33-fold serial dilutions of standard. Transfer 150 μ l from the 25,000pg/ml initial human IFN- β dilution (Step 3) to 350 μ l of culture medium for the second dilution. Mix by pipetting and repeat five times to generate seven standard dilutions with a range of 25,000pg/ml to 18.2pg/ml. The last well or chamber should contain only culture medium as the background control.

Note: Change pipette tips between each dilution step and use aerosol filter tips to avoid analyte carryover.

6. After the cell treatment is complete, add the standard dilutions and background control in duplicate to two columns in the assay plate (see the plate map in Section 3).

96-well plate: Dispense 80 μ l per well.

384-well plate: Dispense 20 μ l per well.

Notes:

- a. Unused Human IFN- β Standard (10 μ g/ml) can be stored at 4 $^{\circ}$ C for 1 month. If storing the standard for more than 1 month, dispense into aliquots and store at less than -65 $^{\circ}$ C. Avoid multiple freeze-thaw cycles.
- b. We recommend incorporating Human IFN- β Standard controls on each assay plate for normalization.
- c. Purchase additional Human IFN- β Standard (10 μ g/ml; Cat.# W149A-C) if needed.

4.C. Adding 5X Anti-hIFN- β Antibody Mixture to Assay Wells

If using multiple cell models, each requiring a different culture medium, separate 5X Anti-hIFN- β antibody mixtures must be generated in each medium used in the study.

1. Remove the Anti-hIFN- β antibodies from storage immediately before use. Thaw if necessary.
Note: Remove Lumit[®] Detection Buffer B from storage at the same time and equilibrate to room temperature if not already thawed. Use a water bath to accelerate buffer warming as necessary. If using the 25ml buffer in Cat.# W1811, you may need to initiate buffer warming further in advance of its use in Section 4.D.
2. Briefly centrifuge the Anti-hIFN- β antibody tubes before opening, then mix by pipetting.
3. Immediately prior to use, prepare a 5X antibody mixture by diluting both antibodies 1:200 into a single volume of prewarmed culture medium. Pipette to mix the antibody solution. To assay a complete 96- or 384-well plate, including some reagent volume for pipetting loss, prepare the 5X antibody mixture as follows:

| Reagent | Volume |
|------------------------------|---------------|
| culture medium | 2,376 μ l |
| Anti-hIFN- β mAb-SmBiT | 12 μ l |
| Anti-hIFN- β mAb-LgBiT | 12 μ l |

4. Add the 5X Anti-hIFN- β antibody mixture to wells containing cultured cells or standard dilutions, carefully avoiding cross contamination between wells by changing pipette tips if moving from high to low analyte levels.
96-well plate: Dispense 20 μ l/well of 5X Anti-hIFN- β antibody mixture to 80 μ l/well of cells or IFN- β standard dilutions.
384-well plate: Dispense 5 μ l/well of 5X Anti-hIFN- β antibody mixture to 20 μ l/well of cells or IFN- β standard dilutions.
5. Briefly mix with a plate shaker (e.g., 10 seconds at 250–350rpm).
6. Incubate for 45 minutes at 37°C in a humidified 5% CO₂ incubator.

Notes:

- a. This incubation step can be performed at room temperature and ambient atmosphere if the effect on cell responses and cell health is not a concern.
- b. To incubate at room temperature, a HEPES-containing medium will provide best results. Without HEPES, the buffering capacity outside of a CO₂ incubator is limited. The plates can also be incubated at 37°C in a CO₂ incubator if subsequently equilibrated to room temperature prior to adding detection reagent.

4.D. Adding Lumit[®] Detection Reagent B to Assay Wells

While cells are incubating with the Anti-hIFN- β antibody mixture (Section 4.C), prepare the Lumit[®] detection reagent B.

1. Equilibrate the required volume of Lumit[®] Detection Buffer B to room temperature.
2. Remove the Lumit[®] Detection Substrate B from storage and mix. If Lumit[®] Detection Substrate B has collected in the cap or on the sides of the tube, briefly centrifuge.
3. Prepare a 1:20 dilution of Lumit[®] Detection Substrate B in room temperature Lumit[®] Detection Buffer B to create enough volume of Lumit[®] detection reagent B for the number of wells to be assayed. To assay a 96- or 384-well assay plate, including some reagent volume for pipetting loss, prepare 5X Lumit[®] detection reagent B as follows:

| Reagent | Volume |
|--|---------------|
| Lumit [®] Detection Buffer B | 3,040 μ l |
| Lumit [®] Detection Substrate B | 160 μ l |

Notes:

- a. The 1,000-assay size Lumit[®] IFN- β (Human) Immunoassay (Cat.# W1811) contains 25ml of Lumit[®] Detection Buffer B and 1.25ml of Lumit[®] Detection Substrate B. There is sufficient volume to prepare Lumit[®] detection reagent B for analyzing 5 or 10 plates at one time. If Cat.# W1811 is used to assay 10 plates individually, mix 2,375 μ l of Lumit[®] Detection Buffer B + 125 μ l of Lumit[®] Detection Substrate B for each plate.
 - b. Once reconstituted, the Lumit[®] detection reagent B will lose 10% activity in approximately 3 hours at 20°C. At 4°C, the reconstituted reagent will lose 10% activity in approximately 7 hours.
4. After the incubation in Section 4.C, Step 6, equilibrate the assay plate with cells to room temperature for 15 minutes.
 5. Add room temperature 5X Lumit[®] detection reagent B to each assay well of the plate:
96-well plate: Dispense 25 μ l per well.
384-well plate: Dispense 6.25 μ l per well.
 6. Briefly mix with a plate shaker (e.g., 10 seconds at 300–500rpm).
 7. Incubate at room temperature for 3–5 minutes.
 8. Read luminescence.

Notes:

- a. Assay signal is stable with a half-life of approximately 2 hours, compatible with batch processing of multiple assay plates. We recommend incorporating the Human IFN- β Standard controls on each assay plate for normalization.
- b. All standard luminescence plate readers are suitable for this assay. An integration time of 0.25–1 second per well should be suitable. The gain settings on some instruments might require optimizing to achieve sensitivity and dynamic range. Consult the instrument manual. The GloMax[®] Discover System (Cat.# GM3000) provides a pre-installed “Lumit Immunoassay” protocol under the ‘Luminescence Protocols’ tab with a 0.5 second integration time.

5. Optional Sample Transfer Protocol

This protocol describes sample medium transfer from treated cell wells into a separate assay plate for subsequent cytokine detection, leaving the cells and remaining medium for additional uses. For quantitation purposes, a dilution series of Human IFN- β Standard prepared in culture medium is used to generate a standard curve.

5.A. Cell Plating and Treatment

1. Plate cells into a 96- or 384-well tissue culture plate. Leave columns 1 and 2 empty. Allow cells to attach if using adherent cells.

Note: While the broad dynamic range of the assay offers considerable flexibility for cell number, the optimal dispensed cell number per well for a specific cell model should be empirically determined. Ensure that the maximum cytokine level released does not exceed the linear range of the detection chemistry. Within that constraint, cell number can be increased to meet the detection requirements in cases of low-level cytokine production.

2. Treat cells by adding a volume of test agent to each well. The final treatment volume is flexible, and can be adjusted to accommodate intended sample analyses, including split-sample analysis for multiple cytokines. Typical volumes are:

96-well plate: 100–200 μ l per well

384-well plate: 25–50 μ l per well

Optional: If manually dispensing into a 384-well assay format, after various additions, briefly centrifuge the microplate (700–900rpm for approximately 10 seconds) then briefly mix with a plate shaker.

3. After cell treatment is complete, transfer cell medium from each well to the corresponding wells of a separate white assay plate:

96-well plate: Transfer 50 μ l per well.

384-well plate: Transfer 12.5 μ l per well.

Notes:

- a. If lower sample volumes are transferred, dilute with culture medium to 50 μ l for 96-well format and 12.5 μ l for 384-well format. Be sure to account for sample dilution when calculating actual cytokine concentration released in cell wells.
- b. Assay sensitivity may be reduced in 384-well format compared to that in 96-well format.
- c. For enhanced 384-well assay sensitivity, if needed, you can transfer 20 μ l of sample (and standards) to assay wells, then add 5 μ l/well of 5X Anti-hIFN- β antibody mixture (see Section 4.C for preparing the more concentrated Anti-hIFN- β antibody mixture).

5.B. Preparing Human IFN- β Standard Dilutions

Shortly before completing cell treatments, prepare IFN- β dilutions in the identical culture medium used for cell samples.

Note: If using multiple cell models, each requiring a different culture medium, separate standard dilution series must be generated in each medium used.

1. Thaw the Human IFN- β Standard (approximately 15 minutes at room temperature) immediately before use.
2. Briefly centrifuge the tube before opening, then mix by pipetting.
3. Prepare an initial concentration of 25,000pg/ml human IFN- β by diluting the Human IFN- β Standard (10 μ g/ml) 1:400 in prewarmed cell culture medium. For example, prepare 800 μ l of 25,000pg/ml human IFN- β by adding 2 μ l of the Human IFN- β Standard stock to 798 μ l of culture medium (Figure 3).
4. Set up seven tubes (or seven chambers in a dilution reservoir) with 350 μ l of culture medium in each.
5. Proceed with 3.33-fold serial dilutions of standard. Transfer 150 μ l from the 25,000pg/ml stock to 350 μ l of culture medium for the second dilution. Mix and repeat five times to generate seven standard dilutions with a range of 25,000pg/ml to 18.2pg/ml. The last well or chamber should contain only culture medium as the background control.

Note: Change pipette tips between each dilution step and use aerosol filter tips to avoid analyte carryover.

6. After transferring the culture medium from the treated cell wells to a separate assay plate, add the standard dilutions and background control in duplicate to two columns in the transfer plate (see the plate map in Section 3).

96-well plate: Dispense 50 μ l per well.

384-well plate: Dispense 12.5 μ l per well.

Notes:

- a. Unused Human IFN- β Standard (10 μ g/ml) can be stored at 4°C for 1 month. If storing the standard for more than 1 month, dispense into aliquots and store at less than -65°C. Avoid multiple freeze-thaw cycles.
- b. We recommend incorporating Human IFN- β Standard controls on each assay plate for normalization.
- c. Purchase additional Human IFN- β Standard (10 μ g/ml; Cat. # W149A-C) if needed.

5.C. Adding 2X Anti-hIFN- β Antibody Mixture to Sample Wells

If using multiple cell models, each requiring a different culture medium, separate 2X Anti-hIFN- β antibody mixtures must be generated in each medium used in the study.

1. Remove the Anti-hIFN- β antibodies from storage immediately before use. Thaw if necessary.
Note: Remove Lumit[®] Detection Buffer B from storage at the same time and equilibrate to room temperature if not already thawed. Use a water bath to accelerate buffer warming as necessary. If using the 25ml buffer in Cat.# W1811, you may need to initiate buffer warming further in advance of its use in Section 5.D.
2. Briefly centrifuge the Anti-hIFN- β antibody tubes before opening, then mix by pipetting.
3. Immediately prior to use, prepare a 2X antibody mixture by diluting both antibodies 1:500 into a single volume of prewarmed cell culture medium. Pipette to mix the antibody solution. To assay a complete 96- or 384-well plate, including some reagent volume for pipetting loss, prepare the 2X antibody mixture as follows:

| Reagent | Volume |
|------------------------------|---------------|
| culture medium | 5,976 μ l |
| Anti-hIFN- β mAb-SmBiT | 12 μ l |
| Anti-hIFN- β mAb-LgBiT | 12 μ l |

4. Add the 2X Anti-hIFN- β antibody mixture to transferred culture medium (samples) or IFN- β standard dilutions, carefully avoiding cross contamination between wells by changing pipette tips if moving from high to low analyte levels.
96-well plate: Dispense 50 μ l/well of 2X Anti-hIFN- β antibody mixture to 50 μ l/well of medium or standard dilutions.
384-well plate: Dispense 12.5 μ l/well of 2X Anti-hIFN- β antibody mixture to 12.5 μ l/well of medium or standard dilutions.
5. Briefly mix with a plate shaker (e.g., 10 seconds at 250–350rpm).
6. Incubate for 60 minutes at room temperature.
Note: To incubate at room temperature, a HEPES-containing medium will provide best results. Without HEPES, the buffering capacity outside of a CO₂ incubator is limited. The plates can also be incubated at 37°C in a CO₂ incubator if subsequently equilibrated to room temperature prior to adding detection reagent.

5.D. Adding Lumit[®] Detection Reagent B to Sample Wells

While the samples and standard dilutions are incubating with the Anti-hIFN- β antibody mixture (Section 5.C), prepare the Lumit[®] detection reagent B.

1. Equilibrate the required volume of Lumit[®] Detection Buffer B to room temperature.
2. Remove the Lumit[®] Detection Substrate B from storage and mix. If the Lumit[®] Detection Substrate B has collected in the cap or on the sides of the tube, briefly centrifuge.
3. Prepare a 1:20 dilution of Lumit[®] Detection Substrate B into room temperature Lumit[®] Detection Buffer B to create enough volume of Lumit[®] detection reagent B for the number of wells to be assayed. For a 96- or 384-well assay plate, including some excess reagent volume, prepare 5X Lumit[®] detection reagent B as follows:

| Reagent | Volume |
|--|---------------|
| Lumit [®] Detection Buffer B | 3,040 μ l |
| Lumit [®] Detection Substrate B | 160 μ l |

Notes:

- a. The 1,000-assay size of Lumit[®] IFN- β (Human) Immunoassay (Cat.# W1811) contains 25ml of Lumit[®] Detection Buffer B and 1.25ml of Lumit[®] Detection Substrate B. There is sufficient volume to prepare Lumit[®] detection reagent B for analyzing 5 or 10 plates at once. If Cat.# W1811 is used for assaying 10 plates individually, mix 2,375 μ l of Lumit[®] Detection Buffer B + 125 μ l of Lumit[®] Detection Substrate B for each plate.
 - b. Once reconstituted, the Lumit[®] detection reagent B loses 10% activity in approximately 3 hours at 20°C. At 4°C, the reconstituted reagent loses 10% activity in approximately 7 hours.
4. After the incubation in Section 5.C, Step 6, add room temperature Lumit[®] detection reagent B to each assay well of the plate:
96-well plate: Dispense 25 μ l per well.
384-well plate: Dispense 6.25 μ l per well.
 5. Briefly mix with a plate shaker (e.g., 10 seconds at 300–500rpm).
 6. Incubate at room temperature for 3–5 minutes.
 7. Read luminescence.

Notes:

- a. Assay signal is stable with a half-life of approximately 2 hours, compatible with batch processing of multiple assay plates.
- b. All standard plate readers capable of reading luminescence are suitable for this assay. An integration time of 0.25–1 second per well should be suitable. Some instruments might require optimizing the gain settings to achieve sensitivity and dynamic range. Consult the instrument manual for instrument settings. The GloMax[®] Discover System (Cat.# GM3000) provides a pre-installed “Lumit Immunoassay” protocol under the ‘Luminescence Protocols’ tab with a 0.5 second integration time.

6. Calculating Results

Create a standard curve for the known cytokine concentrations using software (e.g., GraphPad® Prism) capable of nonlinear regression analysis or cubic spline curve fitting.

Subsequently, interpolate the concentration of cytokine in various cell samples. The broad dynamic range of the Lumit® standard curve closely approaches linearity and is well-suited for second- or third-order polynomial regression curve fitting, as well as cubic spline curve fitting. Four-parameter logistic (4PL) curve fitting is also commonly used, but may not be ideal since the broad, linear dynamic range for the Lumit® standard curve is not well-suited for sigmoidal curve fitting (3).

Alternatively, while somewhat less accurate, a Log-Log plot of average relative light units (RLU; background-subtracted) vs. cytokine standard concentrations can be fit with the Power trendline in Microsoft Excel® (see Section 7, Figure 4) and subsequently used for interpolation of the concentration of cytokine release in various cell samples.

7. Representative Data

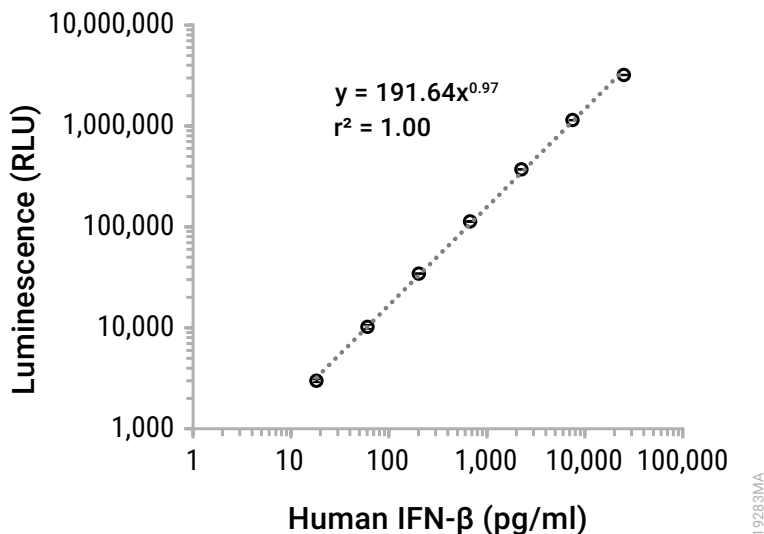


Figure 4. Standard curve for the Lumit® IFN-β (Human) Immunoassay. This is a representative standard curve and should not be used for calculation of unknowns. Generate a standard curve on each assay plate to interpolate the concentration of cytokine in experimental samples. **Note:** The plotted luminescence values were determined by subtracting background RLU.

Table 1. Intra-Assay Precision. Three samples of known concentrations of human IFN- β were tested with 20 replicates on one plate to assess intra-assay precision. A standard curve was used on the assay plate to interpolate the IFN- β quantities in each well by the various methods using GraphPad® Prism software.

| | Cubic spline fit | | | Third order polynomial (cubic) fit | | |
|-----------------------------|------------------|--------|--------|------------------------------------|--------|-------|
| | 20 | 20 | 20 | 20 | 20 | 20 |
| n | 20 | 20 | 20 | 20 | 20 | 20 |
| Expected (pg/ml) | 20,000 | 2,000 | 200 | 20,000 | 2,000 | 200 |
| Mean (pg/ml) | 20,160 | 2,026 | 195 | 20,596 | 2,033 | 189 |
| Standard Deviation | 802 | 77 | 6.1 | 770 | 72 | 5.7 |
| Percent CV | 4.0 | 3.8 | 3.1 | 3.7 | 3.5 | 3.0 |
| Average Percent of Expected | 100.8 | 101.3 | 97.7 | 103.0 | 101.6 | 94.5 |
| Percent Range | 95–107 | 94–109 | 92–103 | 97–109 | 94–109 | 89–99 |

Note: The minimal detectable dose (MDD), determined at two standard deviations above background, was 4.6pg/ml.

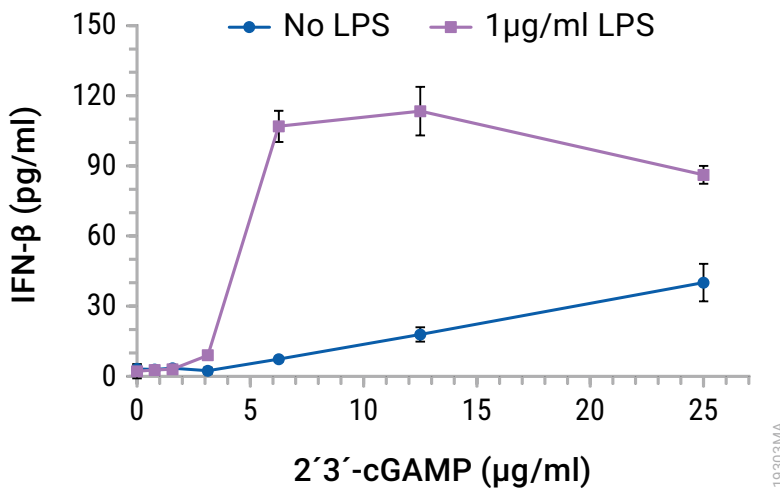


Figure 5. Lumit® detection of IFN- β released from human THP-1 cells. THP-1 cells were plated in 96-well plate format at 50,000 cells/well in RPMI 1640 + 10% heat-inactivated FBS. Cells were treated in triplicate for 40 hours with increasing concentrations of 2'3'-cGAMP (InvivoGen Cat.# tlr-nacga23s) with and without adding 1 μ g/ml lipopolysaccharide (LPS; InvivoGen. Cat.# tlr-b5lps) for the last 20 hours of treatment. Following treatment, the 5X Anti-hIFN- β antibody mixture was dispensed into the cell wells and incubated for 1 hour before Lumit® detection reagent B was added. Luminescence readings for unknowns were interpolated against a standard curve using cubic spline fitting (GraphPad® Prism) to determine levels of IFN- β released.

8. Troubleshooting

For questions not addressed here, please contact your local Promega Branch Office or Distributor. Contact information available at: www.promega.com. Email: techserv@promega.com

| Symptoms | Causes and Comments |
|--|--|
| No signal from treated cells | Use only human cells with the Lumit [®] IFN- β (Human) Immunoassay. The Lumit [®] IFN- β (Human) Immunoassay will not effectively detect IFN- β from other species. In addition to the Human IFN- β Standard, consider using treated human THP-1 cells as a positive control. |
| Human IFN- β standard curve is not linear | The 3.33-fold dilutions should be carefully created without carryover from a higher concentration. We recommend changing aerosol filter pipette tips after each dilution step to prevent carryover. The assay sensitivity and the broad linear range (>3 logs) means that any carryover will disrupt the linear range. Also, make sure that no IFN- β contaminates the background control. |
| The relative light units (RLU) for the standard are low and/or variable | Warm culture medium, Human IFN- β Standard and Lumit [®] Detection Buffer B to room temperature before use. There may be some variation in RLU due to culture conditions, temperature, etc., but as long as the standard curve is run on the same plate as the test samples under the same conditions, released IFN- β can be accurately quantitated. |
| High assay background and reduced signal-to-background ratio when using medium with low or no FBS (or albumin) | Using medium with less than 5% FBS can produce higher background (nonanalyte-mediated antibody pairing) and reduce assay sensitivity. When using serum-free, albumin-free medium, you can reduce assay background and improve sensitivity by preparing the assay antibody mixture in medium supplemented with BSA to deliver 0.05% BSA final after addition to sample. (Note: Using significantly higher albumin concentrations than indicated can further compromise assay performance.) |
| Greatly reduced assay sensitivity when applied to human serum samples | Several factors within human serum samples can interfere with the homogeneous Lumit [®] IFN- β (Human) Immunoassay performance and, thereby, significantly reduce assay sensitivity. For this reason, we do not recommend use of the Lumit [®] IFN- β (Human) Immunoassay on human serum samples. |

Symptoms

Experimental RLU values are not identical to the numbers in this Technical Manual

Causes and Comments

Bioluminescent signal intensity (i.e., absolute RLU) will vary between laboratories due to several factors, such as specific experimental conditions (buffers, volumes), as well as plates and plate readers. Establish the assay performance in your lab with the sample buffer, plates and instruments you will be using. The Human IFN- β Standard provided in the kit can be used to generate a standard curve that will help establish the parameters expected in your lab.

9. Appendix**9.A. References**

1. Dixon, A.S. *et al.* (2016) NanoLuc complementation reporter optimized for accurate measurement of protein interactions in cells. *ACS Chem. Biol.* **11**, 400–8.
2. Hwang, B.B. *et al.* (2020) A homogeneous bioluminescent immunoassay to probe cellular signaling pathway regulation. *Commun. Biol.* **3**, 8.
3. Herman, R.A., Scherer, P.N. and Shan, G. (2008) Evaluation of logistic and polynomial models for fitting sandwich-ELISA calibration curves. *J. Immunol. Methods* **339**, 245–58.

9.B. Related Products

Lumit[®] Cytokine Immunoassays and Supporting Reagents

| Product | Size | Cat.# |
|--|------------|---------|
| Lumit [®] Human IFN- β Standard | 25 μ l | W149A-C |

Additional cytokine standards available.

| Product | Size | Cat.# |
|--|------------|-------|
| Lumit [®] HMGB1 (Human/Mouse) Immunoassay | 100 assays | W6110 |
| Lumit [®] IFN- γ (Human) Immunoassay | 100 assays | W6040 |
| Lumit [®] IL-1 β (Human) Immunoassay | 100 assays | W6010 |
| Lumit [®] IL-1 β (Mouse) Immunoassay | 100 assays | W7010 |
| Lumit [®] IL-2 (Human) Immunoassay | 100 assays | W6020 |
| Lumit [®] IL-4 (Human) Immunoassay | 100 assays | W6060 |
| Lumit [®] IL-6 (Human) Immunoassay | 100 assays | W6030 |
| Lumit [®] IL-10 (Human) Immunoassay | 100 assays | W6070 |
| Lumit [®] IL-17A (Human) Immunoassay | 100 assays | W1430 |
| Lumit [®] TNF- α (Human) Immunoassay | 100 assays | W6050 |

Additional sizes available.

| Product | Size | Cat.# |
|---|------------|--------|
| Lumit Detection Reagent B* | 100 assays | VB4050 |
| Lumit [®] Immunoassay Cellular Systems–Starter Kit | 200 assays | W1220 |
| Lumit [®] Immunoassay Labeling Kit* | 1 each | VB2500 |

*Additional sizes available.

Inflammation Assays

| Product | Size | Cat.# |
|---|------------|--------|
| Caspase-Glo [®] 1 Inflammasome Assay | 10ml | G9951 |
| RealTime-Glo [™] Extracellular ATP Assay | 200 assays | GA5010 |

Additional sizes available.

Cell Viability Assays

| Product | Size | Cat.# |
|---|---------------|--------------|
| CellTiter-Glo [®] 2.0 Cell Viability Assay | 10ml | G9241 |
| RealTime-Glo [™] MT Cell Viability Assay | 100 reactions | G9711 |
| CellTiter-Fluor [™] Cell Viability Assay | 10ml | G6080 |

Additional sizes available.

Cytotoxicity Assays

| Product | Size | Cat.# |
|---|-------------|--------------|
| LDH-Glo [™] Cytotoxicity Assay | 10ml | J2380 |
| CytoTox-Glo [™] Cytotoxicity Assay | 10ml | G9290 |
| CellTox [™] Green Cytotoxicity Assay | 10ml | G8741 |

Additional sizes available.

Apoptosis Assays

| Product | Size | Cat.# |
|--|-------------|--------------|
| Caspase-Glo [®] 3/7 Assay System | 2.5ml | G8090 |
| Caspase-Glo [®] 8 Assay System | 2.5ml | G8200 |
| Caspase-Glo [®] 9 Assay System | 2.5ml | G8210 |
| RealTime-Glo [™] Annexin V Apoptosis and Necrosis Assay | 100 assays | JA1011 |

Additional sizes available.



9.B. Related Products (continued)

Energy Metabolism and Oxidative Stress Assays

| Product | Size | Cat.# |
|--|------|--------|
| NAD/NADH-Glo™ Assay | 10ml | G9071 |
| NADP/NADPH-Glo™ Assay | 10ml | G9081 |
| ROS-Glo™ H ₂ O ₂ Assay | 10ml | G8820 |
| GSH/GSSG-Glo™ Assay | 10ml | V6611 |
| Lactate-Glo™ Assay | 5ml | J5021 |
| Glucose-Glo™ Assay | 5ml | J6021 |
| Glucose Uptake-Glo™ Assay | 5ml | J1341 |
| Glycogen-Glo™ Assay | 5ml | J5051 |
| Pyruvate-Glo™ Assay | 5ml | J4051 |
| Malate-Glo™ Assay | 5ml | JE9400 |
| Glutamine/Glutamate-Glo™ Assay | 5ml | J8021 |
| Glycerol-Glo™ Assay | 5ml | J3150 |
| Triglyceride-Glo™ Assay | 5ml | J3160 |
| Cholesterol/Cholesterol Ester-Glo™ Assay | 5ml | J3190 |
| BHB-Glo™ (Ketone Body) Assay | 5ml | JE9500 |
| BCAA-Glo™ Assay | 5ml | JE9300 |

Additional sizes available.

^(a)U.S. Pat. Nos. 9,797,889; 9,797,890; 10,107,800; 10,648,971; and other patents and patents pending.

^(b)U.S. Pat. No. 8,809,529, European Pat. No. 2635582, and other patents and patents pending.

^(c)U.S. Pat. No. 11,493,504; European Pat. Nos. 3783011 and 2970412; and Japanese Pat. No. 7280842.

© 2024 Promega Corporation. All Rights Reserved.

Caspase-Glo, CellTiter-Glo, Lumit, NanoBiT and NanoLuc are registered trademarks of Promega Corporation. CellTiter-Fluor, CellTox, CytoTox-Glo, Ester-Glo, Glucose-Glo, Glutamine/Glutamate-Glo, Glycerol-Glo, GSH/GSSG-Glo, Lactate-Glo, LDH-Glo, NAD/NADH-Glo, NADP/NADPH-Glo, RealTime-Glo, ROS-Glo and Triglyceride-Glo are trademarks of Promega Corporation.

Corning is a registered trademark of Corning, Inc. Dilux is a registered trademark of Dilux, L.L.C. Excel and Microsoft are registered trademarks of Microsoft Corporation. GIBCO is a registered trademark of Life Technologies Corporation. GraphPad Prism is a registered trademark of GraphPad Software, Inc.

Products may be covered by pending or issued patents or may have certain limitations. Please visit our website for more information.

All prices and specifications are subject to change without prior notice.

Product claims are subject to change. Please contact Promega Technical Services or access the Promega online catalog for the most up-to-date information on Promega products.